

PPAR α Transcription Factor Assay Kit

Item No. 10006915

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GENERAL INFORMATION

Materials Supplied

Kit components may be stored at -20°C prior to use. For long term storage, the Positive Control should be thawed on ice, aliquoted at 20 µl/vial, and stored at -80°C. After use we recommend each kit component be stored according to the temperature listed below.

Item Number	Item	Quantity	Storage
10006880	Transcription Factor Binding Assay Buffer (4X)	1 vial	4°C
10007472	Transcription Factor Reagent A	1 vial	-20°C
10007441	Transcription Factor PPARα Positive Control	1 vial	-80°C
10006882	Transcription Factor Antibody Binding Buffer (10X)	1 vial	4°C
10007442	Transcription Factor PPARα Primary Antibody	1 vial	-20°C
400062	Wash Buffer Concentrate (400X)	1 vial	4°C
400035	Polysorbate 20	1 vial	RT
10006885	Transcription Factor PPAR Competitor dsDNA	1 vial	-20°C
10006884	Transcription Factor Goat Anti-Rabbit HRP Conjugate	1 vial	-20°C
10006887	Transcription Factor PPAR 96-Well Strip Plate	1 plate	4°C
400012	96-Well Cover Sheet	1 cover	RT
10006888	Transcription Factor Developing Solution	1 vial	4°C
10006889	Transcription Factor Stop Solution	1 vial	4°C

If any of the items listed above are damaged or missing, please contact our Customer Service department at (800) 364-9897 or (734) 975-3999. We cannot accept any returns without prior authorization.



WARNING: This product is for laboratory research use only: not for administration to humans. Not for human or veterinary diagnostic or therapeutic use.

Precautions

Please read these instructions carefully before beginning this assay.

Kit components may be stored at -20°C prior to use. For long term storage, the Positive Control should be thawed on ice, aliquoted at 20 µl/vial and stored at -80°C. After use we recommend each kit component be stored according to the temperatures listed in the booklet.

For research use only. Not for human or diagnostic use.

If You Have Problems

Technical Service Contact Information

Phone: 888-526-5351 (USA and Canada only) or 734-975-3888

Fax: 734-971-3641

E-Mail: techserv@caymanchem.com

Hours: M-F 8:00 AM to 5:30 PM EST

In order for our staff to assist you quickly and efficiently, please be ready to supply the lot number of the kit (found on the outside of the box).

Storage and Stability

This kit will perform as specified if stored as directed and used before the expiration date indicated on the outside of the box.

Materials Needed But Not Supplied

1. A plate reader capable of measuring absorbance at 450 nm
2. Adjustable pipettes and a repeat pipettor
3. A source of UltraPure water; glass Milli-Q or HPLC-grade water are acceptable
4. 300 mM dithiothreitol (DTT)
5. Nuclear Extraction Kit available from Cayman (Item No. 10009277) or buffers for preparation of nuclear extracts (see pages 7-9)

NOTE: The components in each kit lot have been quality assured and warranted in this specific combination only; please do not mix them with components from other lots.

Background

Peroxisome proliferator-activated receptors (PPARs) are ligand-activated transcription factors belonging to the large family of nuclear hormone receptors. There are three subtypes, PPAR α , β/δ , and γ which are expressed at variable levels in different tissues.¹ PPAR α was first identified based on its control of genes encoding peroxisomal fatty acid oxidation enzymes in response to peroxisome proliferators such as fibric acid derivatives. It is expressed predominantly in the liver, heart and kidneys.¹ Upon activation by ligands, such as free fatty acids, lipids and other small molecules, PPAR α forms a heterodimeric complex with RXR α . This ligand-activated complex binds to specific DNA sequences called response elements, allowing transcription of target genes involved in lipid metabolism. PPAR α is activated by a diverse group of small molecules which include the “fibrate” class of synthetic small molecule ligands, clofibrate, fenofibrate, and bezafibrate.² The modulation of PPAR α transcriptional activity by small molecules is under investigation as potential drug targets in developing treatments for several life altering diseases, including atherosclerosis, obesity, and diabetes.^{3,4}

About This Assay

Cayman's PPAR α Transcription Factor Assay is a non-radioactive, sensitive method for detecting specific transcription factor DNA binding activity in nuclear extracts. A 96-well enzyme-linked immunosorbent assay (ELISA) replaces the cumbersome radioactive electrophoretic mobility shift assay (EMSA). A specific double stranded DNA (dsDNA) sequence containing the peroxisome proliferator response element (PPRE) is immobilized onto the bottom of wells of a 96-well plate (see Figure 1, on page 4). PPARs contained in a nuclear extract, bind specifically to the PPRE. PPAR α is detected by addition of specific primary antibody directed against PPAR α . A secondary antibody conjugated to HRP is added to provide a sensitive colorimetric readout at 450 nm. Cayman's PPAR α Transcription Factor Assay detects human PPAR α . It will not cross-react with PPAR γ or PPAR δ .

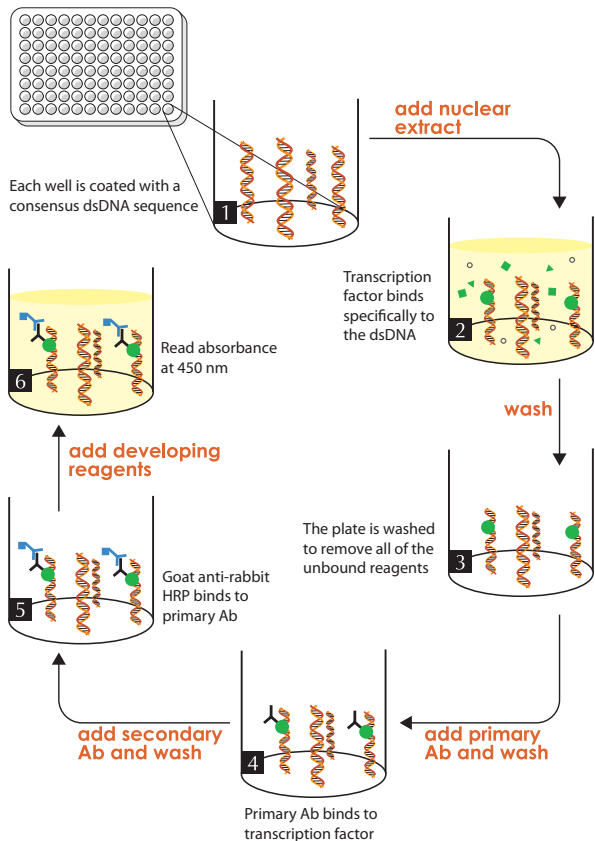


Figure 1. Schematic of the Transcription Factor Binding Assay

PRE-ASSAY PREPARATION

Sample Buffer Preparation

All buffers and reagents below required for preparation of Nuclear Extracts can be purchased directly from Cayman. Catalog numbers for each item are in the Related Products section on page 25. Alternatively, Cayman's Nuclear Extraction Kit (Item No. 10009277) can be used to isolate Nuclear Proteins.

1. PBS (10X)

1.37 M NaCl, 0.027 M KCl, 0.1 M Na_2HPO_4 , 0.017 M KH_2PO_4 , pH 7.4

2. PBS (1X)

Dilute 100 ml of 10X stock with 900 ml distilled H_2O

3. Nuclear Extraction Phosphatase Inhibitor Cocktail (50X)

0.05 M β -glycerophosphate and 1M NaF

0.05 M Na_3VO_4

Store at -80°C

4. PBS/Phosphatase Inhibitor Solution

Add 200 μl of 50X Phosphatase Inhibitor Solution to 10 ml of 1X PBS, mix well, and keep on ice. Make fresh daily.

5. Nuclear Extraction Protease Inhibitor Cocktail (100X)

10 mM AEBSF

0.5 mM Bestatin

0.2 mM Leupeptin Hemisulfate Salt

0.15 mM E-64

0.1 mM Pepstatin A

0.008 mM Aprotinin from Bovine Lung

Made in DMSO, store at -80°C

6. Nuclear Extraction Hypotonic Buffer (10X)

100 mM HEPES, pH 7.5, containing 40 mM NaF, 100 μ M Na₂MoO₄, and 1 mM EDTA

Store at 4°C

7. Complete Extraction Hypotonic Buffer (1X)

Prepare as outlined in Table 1. The phosphatase and protease inhibitors lose activity shortly after dilution; therefore any unused 1X Complete Extraction Hypotonic Buffer should be discarded.

Reagent	150 mm plate ~1.5 x 10 ⁷ cells
Hypotonic Buffer (10X)	100 μ l
Phosphatase Inhibitors (50X)	20 μ l
Protease Inhibitors (100X)	10 μ l
Distilled Water	870 μ l
Total Volume	1,000 μ l

Table 1. Preparation of Complete Extraction Hypotonic Buffer

8. Nonidet P-40 Assay Reagent (10%)

Nonidet P-40 or suitable substitute at a concentration of 10% (v/v) in H₂O

Store at room temperature

9. Nuclear Extraction Buffer (2X)

20 mM HEPES, pH 7.9, containing, 0.2 mM EDTA, 3 mM MgCl₂, 840 mM NaCl, and 20% glycerol (v/v)

Store at 4°C

10. Complete Nuclear Extraction Buffer (1X)

Prepare as outlined in Table 2. Some of the phosphatase and protease inhibitors lose activity shortly after dilution; therefore any remaining 1X Extraction Buffer should be discarded.

Reagent	150 mm plate ~1.5 x 10 ⁷ cells
Nuclear Extraction Buffer (2X)	75 μ l
Protease Inhibitors (100X)	1.5 μ l
Phosphatase Inhibitors (50X)	3.0 μ l
DTT (10 mM)	15 μ l
Distilled Water	55.5 μ l
Total Volume	150 μ l

Table 2. Preparation of Complete Nuclear Extraction Buffer

Purification of Cellular Nuclear Extracts

Cayman's Nuclear Extraction Kit (Item No. 10009277) can be used to isolate nuclear proteins. Alternatively, the procedure described below can be used for a 15 ml cell suspension grown in a T75 flask or adherent cells (100 mm dish 80-90% confluent) where 10^7 cells yields approximately 50 μg of nuclear protein.

1. Collect $\sim 10^7$ cells in pre-chilled 15 ml tubes.
2. Centrifuge suspended cells at 300 x g for five minutes at 4°C.
3. Discard the supernatant. Resuspend cell pellet in 5 ml of ice-cold PBS/Phosphatase Inhibitor Solution and centrifuge at 300 x g for five minutes at 4°C. Repeat one time.
4. Discard the supernatant. Add 500 μl ice-cold 1X Hypotonic buffer. Mix gently by pipetting and transfer resuspended pellet to a pre-chilled 1.5 ml microcentrifuge tube.
5. Incubate cells on ice for 15 minutes allowing cells to swell.
6. Add 100 μl of 10% Nonidet P-40 (or suitable substitute). Mix gently by pipetting.
7. Centrifuge for 30 seconds (pulse spin) at 4°C in a microcentrifuge. Transfer the supernatant which contains the cytosolic fraction to a new tube and store at -80°C.
8. Resuspend the pellet in 50 μl ice-cold Complete Nuclear Extraction Buffer (1X) (with protease and phosphatase inhibitors). Vortex 15 seconds at highest setting then gently rock the tube on ice for 15 minutes using a shaking platform. Vortex sample for 30 seconds at highest setting and gently rock for an additional 15 minutes.
9. Centrifuge at 14,000 x g for 10 minutes at 4°C. The supernatant contains the nuclear fraction. Aliquot to clean chilled tubes, flash freeze and store at -80°C. Avoid freeze/thaw cycles. The extracts are ready to use in the assay.
10. Keep a small aliquot of the nuclear extract to quantitate the protein concentration.

Reagent Preparation

1. Transcription Factor Antibody Binding Buffer (10X)

One vial (Item No. 10006882) contains 3 ml of a 10X stock of Transcription Factor Antibody Binding Buffer (ABB) to be used for diluting the primary and secondary antibodies. To prepare 1X ABB, dilute 1:10 by adding 27 ml of UltraPure water. Store at 4°C for up to six months.

2. Wash Buffer Concentrate (400X)

One vial (Item No. 400062) contains 5 ml of 400X Wash Buffer. Dilute the contents of the vial to a total volume of 2 liters with UltraPure water and add 1 ml of Polysorbate 20 (Item No. 400035). *NOTE: Polysorbate 20 is a viscous liquid and cannot be measured by a pipette. A positive displacement device such as a syringe should be used to deliver small quantities accurately.* A smaller volume of Wash Buffer Concentrate can be prepared by diluting the Wash Buffer Concentrate 1:400 and adding Polysorbate 20 (0.5 ml/liter of Wash Buffer). Store at 4°C for up to two months.

3. Transcription Factor Binding Assay Buffer (4X)

One vial (Item No. 10006880) contains 3 ml of a 4X stock of Transcription Factor Binding Assay Buffer (TFB). Prepare Complete Transcription Factor Binding Assay Buffer (CTFB) immediately prior to use in 1.5 ml centrifuge tubes or 15 ml conical tubes as outlined in Table 3, on page 12. This buffer is now referred to as CTFB. *It is recommended that the CTFB be used the same day it is prepared.*

Component	Volume/Well	Volume/Strip	Volume/ 96-well plate
UltraPure Water	73 μ l	584 μ l	7,008 μ l
4X Transcription Factor Binding Assay Buffer	25 μ l	200 μ l	2,400 μ l
Reagent A (Item No. 10007472)	1 μ l	8 μ l	96 μ l
300 mM DTT	1 μ l	8 μ l	96 μ l
Total Required	100 μ l	800 μ l	9,600 μ l

Table 3. Preparation of Complete Transcription Factor Binding Assay Buffer

4. Transcription Factor PPAR α Positive Control

One vial (Item No. 10007441) contains 150 μ l of clarified cell lysate. This lysate is provided as a positive control for PPAR α activation; it is not intended for plate to plate comparisons. The cell lysate provided is sufficient for 15 reactions and will provide a strong signal (>0.5 AU at 450 nm) when used at 10 μ l/well. When using this positive control, a decrease in signal may occur with repeated freeze/thaw cycles. It is recommended that the Positive Control be aliquoted at 20 μ l per vial and stored at -80°C to avoid loss in signal from repeated freeze/thaw cycles.

ASSAY PROTOCOL

Plate Set Up

There is no specific pattern for using the wells on the plate. A typical layout of PPAR α Positive Control (PC), Competitor dsDNA (C1), and samples of nuclear extracts (S1-S44) to be measured in duplicate is given below in Figure 2. We suggest you record the contents of each well on the template sheet provided (see page 27).

	1	2	3	4	5	6	7	8	9	10	11	12
A	S1	S1	S9	S9	S17	S17	S25	S25	S33	S33	S41	S41
B	S2	S2	S10	S10	S18	S18	S26	S26	S34	S34	S42	S42
C	S3	S3	S11	S11	S19	S19	S27	S27	S35	S35	S43	S43
D	S4	S4	S12	S12	S20	S20	S28	S28	S36	S36	S44	S44
E	S5	S5	S13	S13	S21	S21	S29	S29	S37	S37	NSB	NSB
F	S6	S6	S14	S14	S22	S22	S30	S30	S38	S38	PC	PC
G	S7	S7	S15	S15	S23	S23	S31	S31	S39	S39	Blk	Blk
H	S8	S8	S16	S16	S24	S24	S32	S32	S40	S40	C1	C1

S1-S44 - Sample Wells

NSB - Non-specific Binding Wells

PC - Positive Control Wells

Blk - Blank Wells

C1 - Competitor dsDNA Wells

Figure 2. Sample plate format

- Use the adhesive cover sheet (Item No. 400012) provided to seal the plate.
- Incubate the plate for one hour at room temperature without agitation.
- Empty the wells and wash each well five times with 200 μl of 1X Wash Buffer. After each wash, empty the contents of the plate into the sink. After the final wash (wash #5), tap the plate three to five times on a paper towel to remove any residual Wash Buffer.

Addition of the Transcription Factor Goat Anti-Rabbit HRP Conjugate

- Dilute the Transcription Factor Goat Anti-Rabbit HRP Conjugate (Item No. 10006884) 1:100 in 1X ABB as outlined in Table 5 below. Add 100 μl of diluted secondary antibody to each well except the Blk wells.

Component	Volume/Well	Volume/Strip	Volume/ 96-well plate
1X ABB	99 μl	792 μl	9,504 μl
Goat Anti-Rabbit HRP Conjugate	1 μl	8 μl	96 μl
Total required	100 μl	800 μl	9,600 μl

Table 5. Dilution of Secondary Antibody

- Use the adhesive cover provided to seal the plate.
- Incubate for one hour at room temperature without agitation.
- Empty the wells and wash five times with 200 μl of 1X Wash Buffer. After each wash, empty the contents of the plate into the sink. After the final wash (wash #5), tap the plate three to five times on a paper towel to remove any residual Wash Buffer.

Develop and Read the Plate:

- To each well being used add 100 μl of Transcription Factor Developing Solution (Item No. 10006888), which has been equilibrated to room temperature.
- Incubate the plate for 15 to 45 minutes at room temperature with gentle agitation protected from light. Allow the wells to turn medium to dark blue prior to adding Transcription Factor Stop Solution (Item No. 10006889). (This reaction can be monitored by taking absorbance measurements at 655 nm prior to stopping the reactions; An OD_{655} of 0.4-0.5 yields an OD_{450} of approximately 1). Monitor development of sample wells to ensure adequate color development prior to stopping the reaction. *NOTE: Do not overdevelop; however Positive Control wells may need to overdevelop to allow adequate color development in sample wells.*
- Add 100 μl of Stop Solution per well being used. The solution within the wells will change from blue to yellow after adding the Stop Solution.
- Read absorbance at 450 nm within five minutes of adding the Stop Solution. Blank the plate reader according to the manufacturer's requirements using the blank wells.

Assay Procedure Summary

NOTE: This procedure is provided as a quick reference for experienced users. Follow the detailed procedure when initially performing the assay.

1. Prepare CTFB as described in the Pre-Assay Preparation section, Table 3 on page 12.
2. Add 90 μl CTFB per sample well (80 μl if adding Competitor dsDNA), 100 μl to Blk and NSB wells).
3. Add 10 μl of Competitor dsDNA (optional) to appropriate wells.
4. Add 10 μl of Positive Control to appropriate wells.
5. Add 10 μl of Sample containing PPAR α to appropriate wells.
6. Incubate overnight at 4°C without agitation.
7. Wash each well five times with 200 μl of 1X Wash Buffer.
8. Add 100 μl of diluted PPAR α Primary Antibody per well (except Blk wells).
9. Incubate one hour at room temperature without agitation.
10. Wash each well five times with 200 μl of 1X Wash Buffer.
11. Add 100 μl of diluted Goat Anti-Rabbit HRP Conjugate (except Blk wells).
12. Incubate one hour at room temperature without agitation.
13. Wash each well five times with 200 μl of 1X Wash Buffer.
14. Add 100 μl of Developing Solution per well.
15. Incubate 15 to 45 minutes with gentle agitation.
16. Add 100 μl of Stop Solution per well.
17. Measure the absorbance at 450 nm.

Steps	Reagent	Blk	NSB	PC	C1	S1-S44
1. Add reagents	CTFB	100 μl	100 μl	90 μl	80 μl	90 μl
	Competitor dsDNA				10 μl	
	Positive Control			10 μl	10 μl	
	Samples					10 μl
2. Incubate	Cover plate and incubate overnight at 4°C without agitation					
3. Wash	Wash all wells five times					
4. Add reagents	Primary Antibody		100 μl	100 μl	100 μl	100 μl
5. Incubate	Cover plate and incubate one hour at room temperature without agitation					
6. Wash	Wash all wells five times					
7. Add reagents	Goat Anti-Rabbit HRP Conjugate		100 μl	100 μl	100 μl	100 μl
8. Incubate	Cover plate and incubate one hour at room temperature without agitation					
9. Wash	Wash all wells five times					
10. Add reagents	Developer Solution	100 μl	100 μl	100 μl	100 μl	100 μl
11. Incubate	Monitor development in wells					
12. Add reagents	Stop Solution	100 μl	100 μl	100 μl	100 μl	100 μl
13. Read	Read plate at wavelength of 450 nm					

Table 6. Quick Protocol Guide

Performance Characteristics

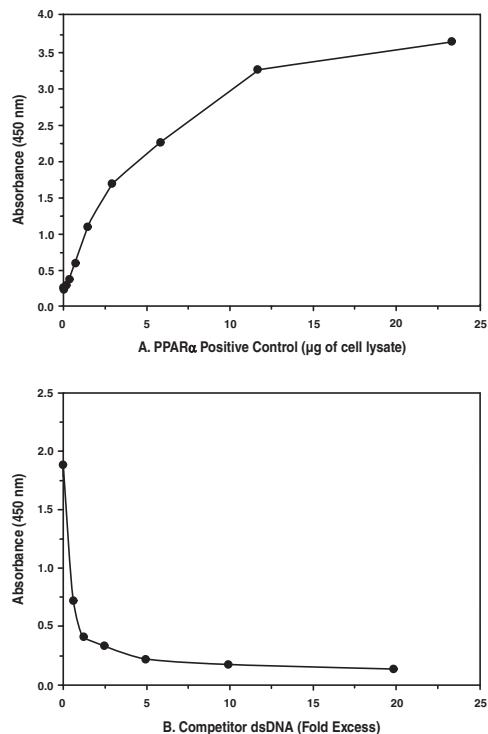


Figure 3. *Panel A:* Increasing amounts of positive control (total lysate) are assayed for PPAR α DNA-binding activity using the Cayman's PPAR α Transcription Factor Assay Kit. *Panel B:* PPAR α DNA-binding assays are performed in the presence of competitive dsDNA. The decrease in signal caused by addition of competitive dsDNA confirms the assay specificity.

Interferences

The following reagents were tested for interference in the assay.

Reagent	Will Interfere (Yes or No)
EGTA (≤ 1 mM)	No
EDTA (≤ 0.5 mM)	No
ZnCl (any concentration)	Yes
DTT (between 1 and 5 mM)	No
Dimethylsulfoxide ($\leq 1.5\%$)	No

Troubleshooting

Problem	Possible Causes	Recommended Solutions
No signal or weak signal in all wells	<ul style="list-style-type: none"> A. Omission of key reagent B. Plate reader settings not correct C. Reagent/reagents expired D. Salt concentrations affected binding between DNA and protein E. Developing reagent used cold F. Developing reagent not added to correct volume 	<ul style="list-style-type: none"> A. Check that all reagents have been added and in the correct order. Perform the assay using the positive control B. Check wavelength setting on plate reader and change to 450 nm C. Check expiration date on reagents D. Reduce the amount of nuclear extract used in the assay, or reduce the amount of salt in the nuclear extracts (alternatively can perform buffer exchange) E. Prewarm the Developing Solution to room temperature prior to use F. Check pipettes to ensure correct amount of developing solution was added to wells
High signal in all wells	<ul style="list-style-type: none"> A. Incorrect dilution of antibody (too high) B. Improper/inadequate washing of wells C. Over-developing 	<ul style="list-style-type: none"> A. Check antibody dilutions and use amounts outlined in instructions B. Follow the protocol for washing wells using the correct number of times and volumes C. Decrease the incubation time when using the developing reagent
High background (NSB)	Incorrect dilution of antibody (too high)	Check antibody dilutions and use amounts outlined in the instructions

Problem (cont.)	Possible Causes (cont.)	Recommended Solutions (cont.)
Weak signal in sample wells	<ul style="list-style-type: none"> A. Sample concentration is too low B. Incorrect dilution of antibody C. Salt concentrations affecting binding between DNA and protein 	<ul style="list-style-type: none"> A. Increase the amount of nuclear extract used. Loss of signal can occur with multiple freeze/thaw cycles of the sample. Prepare fresh nuclear extracts and aliquot as outlined in product insert B. Check antibody dilutions and use amounts outlined in the instructions C. Reduce the amount of nuclear extract used in the assay or reduce the amount of salt in the nuclear extracts (alternatively can perform buffer exchange)

References

1. Olefsky, J.M. Nuclear receptor minireview series. *J. Biol. Chem.* **276(40)**, 36863-36864 (2001).
2. Gervois, P., Torra, I.P., Fruchart, J.-C., *et al.* Regulation of lipid and lipoprotein metabolism by PPAR activators. *Clin. Chem. Lab. Med.* **38(1)**, 3-11 (2000).
3. Park, K.S., Ciaraldi, T.P., Lindgren, K., *et al.* Troglitazone effects on gene expression in human skeletal muscle of type II diabetes involve up-regulation of peroxisome proliferator-activated receptor- γ . *J. Clin. Endocrinol. Metab.* **83**, 2830-2835 (1998).
4. Willson, T.M., Brown, P.J., Sternbach, D.D., *et al.* The PPARs: From orphan receptors to drug discovery. *J. Med. Chem.* **43(4)**, 528-550 (2000).

Related Products

ATF2 (Phospho-Tyr^{69/71}) Transcription Factor Assay Kit - Item No. 600130
ChREBP Transcription Factor Assay Kit - Item No. 10006909
CREB (Phospho-Ser¹³³) Transcription Factor Assay Kit - Item No. 10009846
HIF-1 α Transcription Factor Assay Kit - Item No. 10006910
Liver X Receptor β Transcription Factor Assay Kit - Item No. 10011119
NF- κ B (human p50) Transcription Factor Assay Kit - Item No. 10006912
NF- κ B (human p50/p65) Combo Transcription Factor Assay Kit - Item No. 10011223
NF- κ B p50 (human recombinant) - Item No. 10009818
NF- κ B (p65) Transcription Factor Assay Kit - Item No. 10007889
Nonidet P-40 Assay Reagent (10%) - Item No. 600009
Nuclear Extraction Buffer (2X) - Item No. 10009306
Nuclear Extraction Hypotonic Buffer (10X) - Item No. 10009301
Nuclear Extraction Kit - Item No. 10009277
Nuclear Extraction PBS (10X) - Item No. 10009304
Nuclear Extraction Phosphatase Inhibitors (50X) - Item No. 10009305
Nuclear Extraction Protease Inhibitor Cocktail (100X) - Item No. 10009303
p53 Transcription Factor Assay Kit - Item No. 600020
p53 Designer Transcription Factor Assay Kit - Item No. 600030
PPAR α , δ , γ Complete Transcription Factor Assay Kit - Item No. 10008878
PPAR δ Transcription Factor Assay Kit - Item No. 10006914
PPAR γ Transcription Factor Assay Kit - Item No. 10006855
SREBP-1 Transcription Factor Assay Kit - Item No. 10010854
SREBP-2 Transcription Factor Assay Kit - Item No. 10007819

NOTES

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