PRODUCT INFORMATION



MitoLite[™] Deep Red FX660

Item No. 25160

Ex./Em. Max:	640/659 nm
Supplied as:	A solution in DMSO
Storage:	-20°C
Stability:	≥2 years
Information represents	the product specifications Batch spec

Information represents the product specifications. Batch specific analytical results are provided on each certificate of analysis.

Description

MitoLite[™] reagents are fluorogenic probes that can be used to label mitochondria within live cells. They are hydrophobic and easily cross live cell membranes to accumulate in mitochondria likely via the mitochondrial membrane potential gradient. MitoLite™ reagents contain a cell-retaining group, which allows them to remain in the mitochondria long-term, and their fluorescence can be measured using fluorescence microscopy, microplate fluorometry, or flow cytometry. MitoLite™ Deep Red FX660 displays excitation/emission maxima of 640/659 nm, respectively, and can be used for live and fixed cell applications.

Assay Protocol

1. Prepare mitochondria-staining solution

a. Warm the MitoLite[™] Deep Red FX660 solution to room temperature.

b. Dilute 20 µl of 500X MitoLite™ Deep Red FX660 into 10 ml of Hanks balanced salt solution with 20 mM HEPES (HHBS) or buffer of your choice

Note 1: 20 µl of 500X MitoLite[™] Deep Red FX660 yields a volume of mitochondria-staining solution sufficient for one 96-well plate. Aliquot and store unused MitoLite[™] Deep Red FX660 stock solution at less than -15°C. MitoLite[™] Deep Red FX660 is light sensitive. Light exposure and repeated freeze-thaw cycles should be avoided.

Note 2: The optimal working concentration is application specific. Staining conditions may be modified according to cell type and/or permeability.

2. Prepare and stain the cells

a. Adherent cells:

- i Grow cells in a 96-well black wall/clear bottom plate (100 µl/well) or on coverslips inside a petri dish filled with appropriate culture medium.
- When cells reach the desired confluency, add an equal volume of the mitochondriaii. staining solution prepared in step 1b.
- iii. Incubate cells at 37°C, 5% CO₂ for 30 minutes to 2 hours.
- Wash cells twice with pre-warmed (37°C) HHBS or buffer of your choice, then fill wells iv. with buffer or growth medium.
- v. Observe cells using fluorescence technique of choice.

b. Suspension cells*:

- Centrifuge the cells at 1,000 rpm for 5 minutes to obtain a cell pellet and aspirate the i. supernatant.
- ii. Resuspend cell pellets in pre-warmed (37°C) growth medium and add an equal volume of mitochondria-staining solution prepared in step 1b to cells.
- Incubate cells at 37°C, 5% CO_2 for 30 minutes to 2 hours. iii.
- Wash cells twice with pre-warmed (37°C) HHBS or buffer of your choice, then fill wells iv. with buffer or growth medium.
- Observe cells using fluorescence technique of choice. v.

*Note 3: Suspension cells may be attached to coverslips that have been treated with BD Cell-Tak $^{\circledast}$ (BD Biosciences) and stained as adherent cells.

Note 4: If cells are not sufficiently stained it is recommended to increase either the labeling concentration or the incubation time to increase cellular dye accumulation.

WARNING THIS PRODUCT IS FOR RESEARCH ONLY - NOT FOR HUMAN OR VETERINARY DIAGNOSTIC OR THERAPEUTIC USE.

SAFFTY DATA

This material should be considered hazardous until further information becomes available. Do not ingest, inhale, get in eyes, on skin, or on clothing. Wash thoroughly after handling. Before use, the user must review the complete Safety Data Sheet, which has been sent via email to your institution.

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