TECHNICAL DATA SHEET

THUNDERTM Total c-MET

TR-FRET Cell Signaling Assay Kit

CATALOG NUMBERS KIT-METT-100 (100 tests)

KIT-METT-500 (500 tests)

KIT-METT-2500 (2500 tests)

KIT-METT-5000 (5000 tests)

KIT-METT-10000 (10000 tests)



PRODUCT DESCRIPTION **SPECIFICITY**

This assay kit measures intracellular levels of total c-Met protein in cell lysates using a simple, rapid and sensitive immunoassay based on the homogeneous (no-wash) THUNDER™ TR-FRET technology. The kit is compatible with both adherent and suspension

This assay kit contains two specific and selective antibodies that recognize total (both phosphorylated and unphosphorylated) c-Met.

Store at -80°C

For research use only.

Not for use in diagnostic procedures.

SPECIES REACTIVITY

Human (Swiss-Prot Acc.: P08581; Entrez-Gene Id: 4233).

Other species should be tested on a case-by-case basis.

TR-FRET ASSAY PRINCIPLE

The Total c-Met assay kit is a homogeneous time-resolved Förster resonance energy transfer (TR-FRET) sandwich immunoassay (Figure 1). The THUNDER™ Cell Signaling assay workflow consists of 3 steps (Figure 2). Following cell treatment, cells are first lysed with the specific Lysis Buffer provided in the kit. Then Total c-Met in the cell lysates is detected with a pair of fluorophore-labeled antibodies in a simple "add-incubate-measure" format (single-step reagent addition; no wash steps). One antibody is labeled with a donor fluorophore (Europium chelate: Eu-Abl) and the second with a farred acceptor fluorophore (FR-Ab2). The binding of the two labeled antibodies to distinct epitopes on the target protein takes place in solution and brings the two dyes into close proximity. Excitation of the donor Europium chelate molecules with a flash lamp (320 or 340 nm) or a laser (337 nm) triggers a FRET from the donor to the acceptor molecules, which in turn emit a TR-FRET signal at 665 nm. Residual energy from the Eu chelate generates light at 615 nm. The signal at 665 nm is proportional to the concentration of Total c-Met in the cell lysate. Data can be expressed as either the signal at 665 nm or the 665 nm/615 nm ratio.

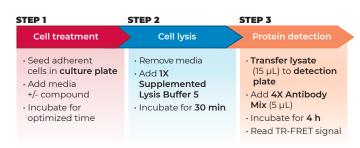


Figure 2 Assay workflow using the 2-plate (transfer) protocol

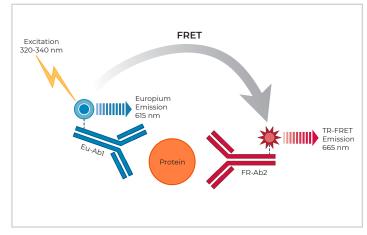


Figure 1 Schematic representation of the TR-FRET cell signaling assay principle.

KIT COMPONENTS

	100 points*	500 points*
Eu-labeled total-c-Met antibody (Eu-Abl)	5 µL	25 μL
Acceptor-labeled total-c-Met antibody (FR-Ab2)	20 μL	100 μL
Lysis Buffer 5 (5X)	1 mL	5 mL
Detection Buffer (10X)	50 μL	250 µL
Positive control cell lysate	100 µL	200 μL
Phosphatase Inhibitor Cocktail (100X)	50 µL	250 µL

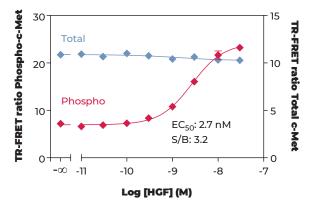
^{*} The number of assay points is based on an assay volume of 20 µL in half-area 96-well or low-volume 384-well assay plates using the kit components at the recommended concentrations (refer to the User Manual).

VALIDATION DATA

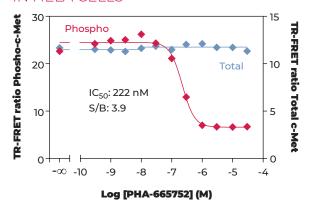
This assay kit has been validated for the relative quantification of total c-Met in HeLa and A431 cell lysates using the 2-plate assay protocol.

- · Adherent cells were cultured overnight in a 96-well tissue culture plate (DMEM +10% FBS).
- Following cell treatment, the media was removed and cells were lysed with the 1X Lysis Buffer 5 (50 µL) supplemented with the phosphatase inhibitors sodium fluoride (1 mM), sodium orthovanadate (2 mM), cOmpleteTM Mini, EDTA-free protease inhibitor cocktail (Roche), bpV(phen) (0.1 mg/mL) and PMSF (1 mM).
- \cdot Following a **30-min** incubation at room temperature (RT) on an orbital shaker (400 rpm), lysates (15 μ L) were then transferred to a 384-well assay plate followed by addition of the labeled antibodies Eu-Ab1 and FR-Ab2 (5 μ L) for detection of total c-Met.
- The plate was incubated at RT for **4 hours** and the TR-FRET signal was recorded at 665 and 615 nm (EnVision®; lamp excitation).

STIMULATION OF PHOSPHO-c-MET (Y1234/Y1235) IN A431 CELLS



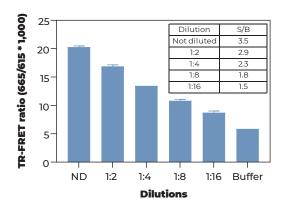
INHIBITION OF PHOSPHO-c-MET (Y1234/Y1235) IN HELA CELLS



A431 cells (50,000 cells/well; in triplicate) were incubated with serial dilutions of of HGF for 10 min at RT. Data show that treatment of A431 cells with HGF stimulates phosphorylation of c-Met at Y1234/Y1235 but does not affect the levels of total c-Met.

HeLa cells (50,000 cells/well; in triplicate) were incubated with serial dilutions of PHA 665752 for 60 min at RT. Cells were then stimulated with 10 nM of HGF for 10 min at RT. Data show that treatment of HeLa cells with PHA-665752 inhibits phosphorylation of c-Met at Y1234/Y1235 by HGF, but does not affect the levels of total c-Met.

HELA CONTROL LYSATE TITRATION (QC TEST)



Quality Control: the Total c-Met assay kit is routinely tested against pervanadate-treated HeLa lysates. HeLa cells were cultured in a T175 flask to 90% confluence and stimulated with 1 mM of pervanadate for 15 min at 37°C. Following cell lysis using 4 mL of 1X Lysis Buffer 5, lysates were serially diluted with 1X Lysis Buffer 5 and tested in triplicate. Data show a linear relationship between lysate dilutions and TR-FRET ratio values.



FOR MORE INFORMATION ON DEVELOPING AND OPTIMIZING TR-FRET CELL SIGNALING ASSAYS, CONSULT THE USER MANUAL.