BRM bromodomain TR-FRET Assay Kit

Item No. 600730

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GENERAL INFORMATION

Materials Supplied

Kit will arrive packaged as a -80°C kit. For best results, remove components and store as stated below.

Item Number	Item	384 wells Quantity/Size	1,920 wells Quantity/Size	9,600 wells Quantity/Size	Storage
600731	BRM bromodomain Europium Chelate	1 vial/ 420 wells	5 vials/ 420 wells	5 vials/ 2,100 wells	-80°C
600732	BRM bromodomain Ligand/APC Acceptor Mixture	1 vial/ 420 wells	5 vials/ 420 wells	5 vials/ 2,100 wells	-80°C
600503	TR-FRET Assay Buffer (10X)	1 vial/2 ml	1 vial/10 ml	5 vials/10 ml	-20°C
600504	TR-FRET Assay Buffer Additive	1 vial/200 mg	1 vial/1 g	5 vials/1 g	-20°C
600506	H3 Positive Control	1 vial/2.5 nmol	5 vials/2.5 nmol	5 vials/12.5 nmol	-80°C
400093	384-Well Solid Plate (low volume; black)	1 plate	5 plates	25 plates	RT
400023	Foil Plate Covers	1 cover	5 covers	25 covers	RT

If any of the items listed above are damaged or missing, please contact our Customer Service department at (800) 364-9897 or (734) 971-3335. We cannot accept any returns without prior authorization.

WARNING: THIS PRODUCT IS FOR RESEARCH ONLY - NOT FOR HUMAN OR VETERINARY DIAGNOSTIC OR THERAPEUTIC USE.

Safety Data

This material should be considered hazardous until further information becomes available. Do not ingest, inhale, get in eyes, on skin, or on clothing. Wash thoroughly after handling.Before use, the user must review the complete Safety Data Sheet, which has been sent *via* email to your institution.

Precautions

Please read these instructions carefully before beginning this assay.

The reagents in this kit have been tested and formulated to work exclusively with Cayman's BRM bromodomain TR-FRET Assay Kit. This kit may not perform as described if any reagent or procedure is replaced or modified.

If You Have Problems

Technical Service Contact Information

Phone: 888-526-5351 (USA and Canada only) or 734-975-3888

Fax: 734-971-3641

Email: techserv@caymanchem.com

Hours: M-F 8:00 AM to 5:30 PM EST

In order for our staff to assist you quickly and efficiently, please be ready to supply the lot number of the kit (found on the outside of the box).

Storage and Stability

This kit will perform as specified in the **Materials Supplied** section, on page 3, and used before the expiration date indicated on the outside of the box.

Materials Needed But Not Supplied

1. A plate reader capable of time-resolved FRET with an excitation at 340 nm and emission at 620 and 670 nm.
2. Adjustable pipettes and a multichannel pipettor.

INTRODUCTION

Background

The acetylation of histone lysine residues plays a crucial role in the epigenetic regulation of gene transcription. Acetylated lysine residues are recognized by a small protein domain known as a bromodomain.[1] These domains recruit regulatory complexes to acetylated nucleosomes, thereby controlling chromatin structure and gene expression. Initial efforts to develop small molecule bromodomain inhibitors have focused on the BET family of proteins, a class of proteins that contain tandem Bromodomains and an Extra Terminal domain.[2] These proteins play a key role in many cellular processes, including inflammatory gene expression, mitosis, and viral/host interactions.[3-5] The isolated individual or tandem bromodomains of many BET family members have been shown to bind acetylated histone tails, serving to couple histone acetylation marks to the transcriptional regulation of target promoters.[4,6-9] Small molecule inhibitors of bromodomain/histone interactions, exemplified by I-BET and JQ-1, hold promise as useful therapeutics for human disease.[10-12]

The SWI/SNF family of ATP-dependent chromatin remodeling enzymes alters the contacts between histones and associated DNA. *In vitro*, these enzymes catalyze structural changes that allow proteins to access the nucleosomal DNA, alter the position of the nucleosomes on the DNA, or eject the histone octamer from the template.[13,14] Mammalian SWI/SNF remodelers are large, multisubunit complexes (>1.5 MDa) that contain either BRM (SMARCA2) or BRG1 (SMARCA4) as their catalytic subunit. In addition to the central snf2 helicase-like domain responsible for the catalytic activity, BRM and BRG1 contain a C-terminal bromodomain that binds acetylated residues on histone tails.[13,14] Stereotypical inhibitors of BET family bromodomains (JQ-1 and I-BET) do not inhibit the interaction of BRM with peptide binding partners (IC_{50} >100 µM). Several subunits of the SWI/SNF complex exhibit tumor suppressor activity and BRM expression is low or absent in several tumor types and its absence correlates with poor prognosis.[15]

About This Assay

Cayman's BRM bromodomain TR-FRET Assay Kit is a homogeneous, Time-Resolved Fluorescence Resonance Energy Transfer (TR-FRET) assay method amenable to rapid characterization of inhibitors of bromodomain/acetylated peptide interaction in a high throughput format. The 'donor' fluorophore in this assay consists of BRM bromodomain (human amino acids 1367-1511) directly labeled with a europium (Eu^{3+}) chelate. A biotinylated peptide containing target acetylated lysine serves as the ligand for the BRM bromodomain. Allophycocyanin (APC)-labeled avidin binds with high affinity to the peptide substrate *via* the biotin moiety and serves as the 'acceptor' fluorophore in the assay. Inhibition of the bromodomain/peptide interaction displaces BRM bromodomain-Eu^{3+} from the APC/avidin resulting in a loss of TR-FRET signal. The BRM bromodomain TR-FRET Assay Kit is robust (Z´ >0.6), and is suitable for high-throughput screening in the provided 384-well plate or can be scaled to higher density plate formats (*e.g.*, 1,536-well plate) if desired. The assay is stable at room temperature for at least four hours and in the presence of less than 2% DMSO.

Introduction to TR-FRET

TR-FRET is based upon the principles of FRET, but possesses a number of advantages that make it a superior technology for high-throughput screening. When an optically active molecule absorbs a photon it has several options by which it may release that energy: it may release a photon of a longer wavelength (less energy) than the photon it absorbed, it may dissipate the energy as heat, or it can transfer the energy non-radiometrically to a suitable acceptor fluorophore. The latter effect is known as FRET and is a commonly used phenomenon in biological assays. In these assays, a donor flurophore is coupled to one binding partner and an acceptor fluorophore is coupled to the other binding partner. The binding partners are mixed in an assay well and allowed to associate. The donor fluorophore is then excited with a wavelength of light that does not excite the acceptor fluorophore and if the molecules are within approximately 100 Å of each other, the donor fluorophore can non-radiometrically transfer the energy to the acceptor fluorophore, which will then release that photon as light with at a wavelength characteristic of the acceptor fluorophore (see Figure 1 on page 9). For each assay point, the fluorescence intensity of the donor fluorophore and the acceptor fluorophore are measured and the data are generally presented as the ratio of acceptor fluorophore intensity/donor fluorophore intensity. This methodology is particularly sensitive because the FRET efficiency decays as the 6th power of the distance between the two fluorophores. Therefore, unassociated binding partners are unlikely to lie within the distance required for efficient FRET.

TR-FRET is an extension of FRET that utilizes a donor fluorophore with a long fluorescent half-life. These fluorophores are based upon lanthanide (most often Eu^{3+} or Tb^{3+}) chelates that have characteristically large stokes shifts and fluorescent half-lives on the order of milliseconds. The long fluorescent lifetime allows the TR-FRET signal to be sustained for dramatically longer periods of time than standard fluorescence. This is particularly advantageous because it affords the ability to measure the TR-FRET signal after background fluorescence in the assay (*e.g.*, buffer/reagent autofluorescence) has dissipated (see Figure 2 on page 10). The increased signal:noise ratio and the diminished effects of screening compound fluorescence makes TR-FRET assays particularly useful for high-throughput screening applications.

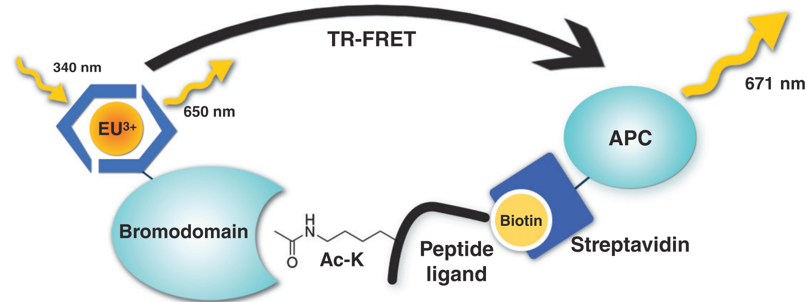

Figure 1. Assay schematic for the bromodomain TR-FRET Assay Kit. Upon excitation, the europium chelate can release a photon or transfer its energy to an APC molecule, provided the APC is in close proximity to the europium fluorophore.

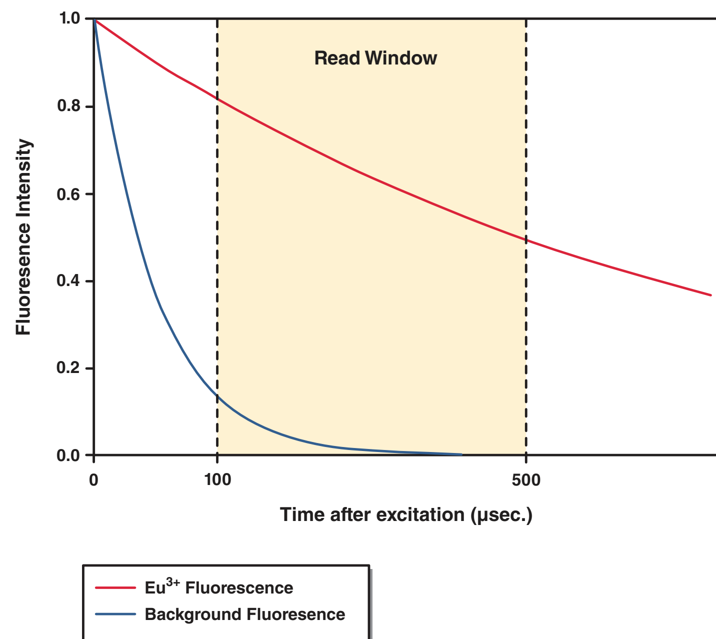

Figure 2. The extended fluorescence lifetimes of Eu^{3+}-based fluorophores allows the samples to be analyzed after background fluorescence has decayed, improving signal to noise and reducing spectral artifacts.

PRE-ASSAY PREPARATION

NOTE: Water used to prepare all reagents and buffers must be deionized and free of trace organic contaminants ('UltraPure'). UltraPure water may be purchased from Cayman Chemical (Item No. 400000).

Buffer Preparation

2 ml vial TR-FRET Assay Buffer (10X) (384-well kit; Item No. 600503): Add 18 ml of UltraPure water to the vial. Add 200 mg of TR-FRET Assay Buffer Additive (Item No. 600504) and mix to dissolve. For best results, filter completed 1X Assay Buffer with a 0.22 µm filter before use. *NOTE: It is normal for the concentrated buffer to contain crystalline salts after thawing. These will completely dissolve upon dilution with water.* Store the diluted buffer at 4°C; it will be stable for approximately one month.

OR

10 ml vial TR-FRET Assay Buffer (10X) (1,920- or 9,600-well kit; Item No. 600503): For five 384-well plates, dilute 10 ml TR-FRET Assay Buffer to a total volume of 100 ml with UltraPure water. Add 1 g of TR-FRET Assay Buffer Additive (Item No. 600504) and mix to dissolve. For best results, filter completed 1X Assay Buffer with a 0.22 µm filter before use. *NOTE: It is normal for the concentrated buffer to contain crystalline salts after thawing. These will completely dissolve upon dilution with water.* Store the diluted buffer at 4°C; it will be stable for approximately one month.

Sample Preparation

All inhibitors, be they small molecules, natural products, or proteins, should be prepared in 1X TR-FRET Assay Buffer at a concentration 4X the desired final assay concentration (*e.g.*, for 1 µM final assay concentration, a 4 µM dilution should be made). This solution may contain up to 8% organic solvents such as DMSO, DMF, or short chain alcohols. The final concentration of organic solvents in the assay will then be <2%. Avoid using high concentrations of metal chelating agents or phosphate buffers.

ASSAY PROTOCOL

Preparation of Assay-Specific Reagents

BRM bromodomain Europium Chelate (Item No. 600731)

420-well vial BRM bromodomain Europium Chelate (384- or 1,920-well kit):

On ice, thaw one tube of BRM bromodomain Europium Chelate (420 wells, Item No. 600731) per 384-well plate and briefly centrifuge before opening. Dilute contents to a final volume of 4.2 ml in 1X TR-FRET Assay Buffer. Content volume is indicated on the label. Mix gently (do not vortex) and keep on ice. Diluted protein should be used within the same day.

OR

2,100-well vial BRM bromodomain Europium Chelate (9,600-well kit):

On ice, thaw one tube of BRM bromodomain Europium Chelate (2,100 wells, Item No. 600731) per five 384-well plates and briefly centrifuge before opening. Dilute contents to a final volume of 21 ml in 1X TR-FRET Assay Buffer. Content volume is indicated on the label. Mix gently (do not vortex) and keep on ice. Diluted protein should be used within the same day.

BRM bromodomain Ligand/APC Acceptor Mixture (Item No. 600732)

420-well vial BRM bromodomain Ligand/APC Acceptor Mixture (384- or 1,920-well kit):

For each 384-well plate, add 2.1 ml of 1X TR-FRET Assay Buffer to one vial of the BRM bromodomain Ligand/APC Acceptor Mixture (420 wells, Item No. 600732) and gently vortex. Keep the solution in the dark to prevent photobleaching. Long-term storage of the diluted mixture is not recommended.

OR

2,100-well vial BRM bromodomain Ligand/APC Acceptor Mixture (9,600-well kit):

For five 384-well plates, add 3 ml of 1X TR-FRET Assay Buffer to one vial of the BRM bromodomain Ligand/APC Acceptor Mixture (2,100 wells, Item No. 600732) and gently vortex. Transfer contents to a new tube and adjust the mixture to a final volume of 10.5 ml in 1X Assay Buffer. Keep the solution in the dark to prevent photobleaching. Long-term storage of the diluted mixture is not recommended.

H3 Positive Control (Item No. 600506)

2.5 nmol vial H3 Positive Control (384- or 1,920-well kit):

For each 384 well plate, add 200 µl of 1X TR-FRET Assay Buffer to one tube containing the H3 Positive Control (2.5 nmol, Item No. 600506) and vortex gently. Unused solutions may be stored at -20°C for approximately two weeks.

OR

12.5 nmol vial H3 Positive Control (9,600-well kit):

For five 384 well plates, add 1 ml of 1X TR-FRET Assay Buffer to one tube containing the H3Positive Control (12.5 nmol, Item No. 600506) and vortex gently. Unused solutions may be stored at -20°C for approximately two weeks.

Performing the Assay

Pipetting Hints
• Use different tips to pipette each reagent.
• Do not expose the pipette tip to the reagent(s) already in the well.
• Avoid introducing bubbles into the wells.

Follow the steps below to accurately measure the TR-FRET ratio in the assay. Allow all reagents except for the BRM bromodomain Europium Chelate to equilibrate to room temperature prior to performing the assay. Keep the BRM bromodomain Europium Chelate on ice until just prior to use. NOTE: Volumes indicated below are for a 384-well plate format with a 20 µl final assay volume. The customer may scale as needed for higher or lower density plate formats.

1. **Inhibitor Samples**

   Dilute inhibitor samples in 1X TR-FRET Assay Buffer to a concentration that is 4X the desired final concentration (*e.g.*, if 1 µM is desired, prepare a 4 µM solution). This solution may contain up to 8% of an organic solvent (*e.g.*, DMSO). Add 5 µl of this solution to the desired wells. For best results, perform the assay in duplicate.

   It is recommended that inhibitor compounds be tested in a concentration-response format with at least eight independent concentrations that span approximately a 1,000-fold range around the expected IC_{50} value of the inhibitor.

2. **Positive and Negative Control Samples**

   For positive (inhibitor control) control wells, add 5 µl of H3 Positive Control to the desired wells. This will provide a final assay concentration of 3 µM unlabeled peptide as a control.

   For negative (no inhibition) control wells, add 5 µl of 1X TR-FRET Assay Buffer to the desired wells. If inhibitor samples from step 1 contain organic solvent, add an equivalent amount of the solvent into the assay in this step.

3. **BRM bromodomain Europium Chelate**

   Add 10 µl of the diluted BRM bromodomain Europium Chelate to every well of the 384-well plate.

4. **Pre-incubation (optional)**

   If desired, incubate the control and sample wells for 15 minutes at room temperature to allow pre-equilibration of the inhibitor and control compounds with the BRM bromodomain Europium Chelate. *Protect from light*.

5. **BRM bromodomain Ligand/APC Acceptor Mixture**

   Add 5 µl of the reconstituted BRM bromodomain Ligand/APC Acceptor Mixture to every well.

6. **Incubation of the Plate**

   Seal the plate with an adhesive aluminum seal and incubate at room temperature for one hour. For automation purposes, the plate does not have to be sealed, but it should remain in the dark to prevent photobleaching.

7. **Reading the Plate**

   Read the plate(s) in time-resolved format by exciting the sample at 340 nm and reading emissions at 620 and 670 nm, using a 100 µs delay and a 500 µs read window. To ensure optimal assay sensitivity, it is strongly recommended that a filter-based instrument be used to perform TR-FRET measurements. The plate reader used at Cayman Chemical employs a 340/40 nm excitation filter, 620/15 nm, and 670/20 nm emission filters. Samples will be stable for analysis for at least five hours if stored at room temperature and protected from light. Data analysis is performed using the TR-FRET ratio (670 nm emission/620 nm emission).

Well Type	H3 Positive Control (µl)	1X Assay Buffer (µl)	Test Sample (µl)	BRM bromodomain Europium Chelate (µl)	BRM bromodomain Ligand/APC Acceptor Mixture (µl)
Positive Control	5	-	-	10	5
Negative Control	-	5*	-	10	5
Experimental Samples	-	-	5	10	5

Table 1. Pipetting summary
*If an organic solvent is used at concentrations >2% in the test samples, include it in the negative control wells at the same concentration as the sample wells to control for solvent effects.

Effects of Solvents

Samples may be prepared in organic solvents such as DMSO, DMF, or short chain alcohols (*e.g.*, MeOH, EtOH), as long as the final concentration of organic solvents in the assay is <2%. High concentrations of metal chelating agents or phosphate buffers may interfere with the fluorescence of the donor fluorophore and should be avoided. If conditions require different solvents or higher concentrations, additional assays may be required to assess solvent interference.

ANALYSIS

Calculations

A plot of the TR-FRET ratio (670 nm emission/620 nm emission) *versus* inhibitor concentration on semi-log axes results in a sigmoidal dose-response curve typical of competitive binding assays. This data can be fit to a 4-parameter logistic equation as shown in Figure 3 on page 19 to calculate IC_{50} values.

Performance Characteristics

Z´ Factor:

Z´ factor is a term used to describe the robustness of an assay, which is calculated using the equation below.[16]

$$Z' = 1 - \frac{3\sigma_{c+} + 3\sigma_{c-}}{|\mu_{c+} - \mu_{c-}|}$$

Where c+: Positive control

c-: Negative control

The theoretical upper limit for the Z´ factor is 1.0. A robust assay has a Z´ factor >0.5. The Z´ factor for Cayman's BRM bromodomain TR-FRET Assay Kit was determined to be 0.62.

Sample Data

The data shown here is an example of the data typically produced with this kit; however, your results will not be identical to these. Do not use the data below to directly compare to your samples. Your results could differ substantially.

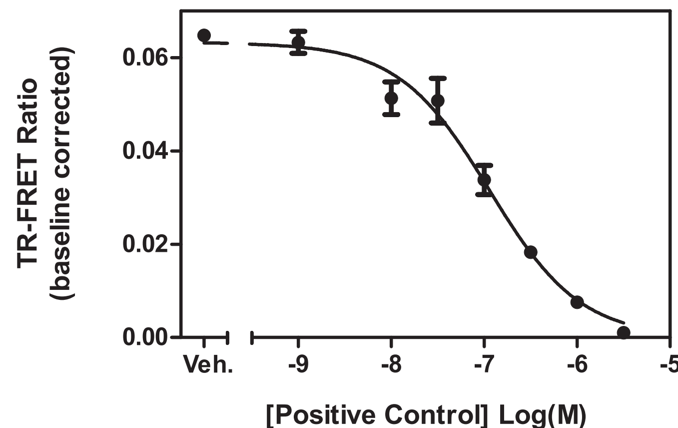

Figure 3. Typical inhibition curves for the displacement of the acetylated peptide from BRM bromodomain by the H3 Positive Control. "Veh." represents compound vehicle control.

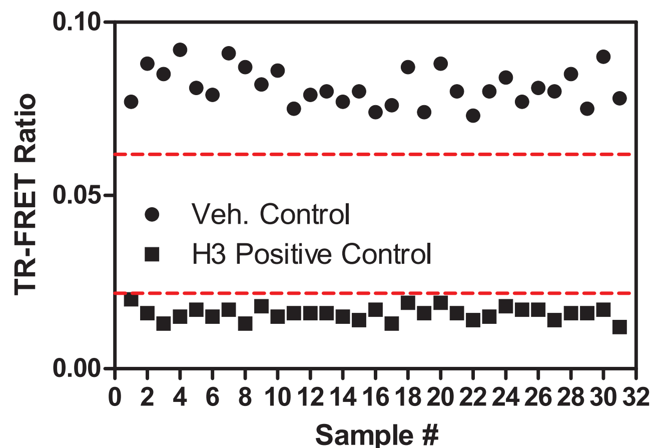

Figure 4. Typical Z´ data for the BRM bromodomain TR-FRET Assay Kit. Data are shown from 64 replicate wells of both positive and negative controls prepared as described in the kit booklet. The calculated Z´ factor from this experiment was 0.62. The red lines correspond to three standard deviations from the mean for each control value.

Troubleshooting

Problem	Possible Causes	Recommended Solutions
Erratic values; dispersion of duplicates	A. Bubble in the well B. Poor pipetting/technique	A. Centrifuge the plate briefly
Low fluorescence signal	A. Incompatible sample matrix B. BRM bromodomain protein handled improperly C. Monochromater-based instrument used for data acqusition	A. Test sample matrix for interference before running samples in the assay B. Keep the protein frozen at -80°C until ready to use; thaw protein and keep on ice until adding to assay C. Analzye the assay using a filter-based plate reader

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NOTES

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