



NOS Activity Assay Kit

Item No. 781001

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GENERAL INFORMATION

Materials Supplied

Kit will arrive packaged as a -80°C kit. After opening kit, store individual components as stated below.

Item Number	Item	Quantity	Storage
10007068	iNOS Positive Control	50 µl	-80°C
10007061	Calmodulin (1 µM)	200 µl	-20°C
10007062	Reaction Buffer (2X)	1.25 ml	-20°C
10007067	L-NNA (10 mM)	100 µl	-20°C
10007063	Homogenization Buffer (10X)	50 ml	RT
10007064	Stop Buffer	25 ml	RT
10007065	Equilibrated Resin	5 ml	RT
10007066	Calcium Chloride (6 mM)	400 µl	RT
601223	Spin Columns and Collection Tubes	50	RT

If any of the items listed above are damaged or missing, please contact our Customer Service department at (800) 364-9897 or (734) 971-3335. We cannot accept any returns without prior authorization.



WARNING: THIS PRODUCT IS FOR RESEARCH ONLY - NOT FOR HUMAN OR VETERINARY DIAGNOSTIC OR THERAPEUTIC USE.

Safety Data

This material should be considered hazardous until further information becomes available. Do not ingest, inhale, get in eyes, on skin, or on clothing. Wash thoroughly after handling. Before use, the user must review the complete Safety Data Sheet, which has been sent *via* email to your institution.

Precautions

Please read these instructions carefully before beginning this assay.

If You Have Problems

Technical Service Contact Information

Phone: 888-526-5351 (USA and Canada only) or 734-975-3888
Fax: 734-971-3641
Email: techserv@caymanchem.com
Hours: M-F 8:00 AM to 5:30 PM EST

In order for our staff to assist you quickly and efficiently, please be ready to supply the lot number of the kit (found on the outside of the box).

Storage and Stability

This kit will perform as specified if stored as listed in the **Materials Supplied** (see page 3) and used before the expiration date indicated on the outside of the box.

Materials Needed But Not Supplied

1. [³H]Arginine monohydrochloride [40-70 Ci/mmol, 1 μCi/μl (Perkin Elmer, Item No. NET1123001MC)] -or- [¹⁴C]arginine [>300 mCi/mmol, 100 μCi/ml (Perkin Elmer, Item No. NEC267E050UC)]
2. Reduced nicotinamide adenine dinucleotide phosphate (NADPH*) (Sigma, Item No. N1630 or N7505)
3. 10 mM Tris-HCl, pH 7.4
4. *Optional:* Magnesium acetate (Sigma, Item No. M2545) for assaying crude iNOS samples (see page 12)
5. Scintillation fluid and vials
6. A source of pure water; glass distilled water or HPLC-grade water is acceptable

*NADPH is not stable in solution; therefore, it is not included in the Reaction Buffer.

About this Assay

Cayman's NOS Activity Assay Kit is a simple, sensitive and specific assay for nitric oxide synthase (NOS) activity. The NOS Activity Assay Kit is based on the biochemical conversion of L-arginine to L-citrulline by NOS.¹⁻⁶ This reaction, which represents a novel enzymatic process, involves a five-electron oxidation of a guanidino nitrogen of L-arginine to nitric oxide (NO), together with the stoichiometric production of L-citrulline (see Figure 1, on page 7). The reaction consumes 1.5 equivalents of reduced nicotinamide adenine dinucleotide phosphate (NADPH) and also requires molecular oxygen, calcium, calmodulin, and tetrahydrobiopterin (BH₄).^{7,8}

Measuring NOS activity by monitoring the conversion of arginine to citrulline is a standard assay for NOS activity in both crude and purified enzyme preparation. Advantages of the NOS Activity Assay Kit include the use of radioactive substrates ([³H]arginine or [¹⁴C]arginine) that enable sensitivity to the picomole level, as well as the specificity of the assay for the NOS pathway due to the direct enzymatic conversion of arginine to citrulline in eukaryotic cells. Additionally, the easy separation of neutrally charged citrulline from positively charged arginine allows multiple assays to be performed easily.

For routine assays, radioactive arginine is added to protein extracts or purified NOS samples. After incubation, the reactions are stopped with a buffer containing ethylenediaminetetraacetic acid (EDTA), which chelates the calcium required by nNOS (neuronal NOS; NOS I) and eNOS (endothelial NOS; NOS III) and, consequently, inactivates the enzyme. In the case of iNOS (inducible NOS; NOS II), the low pH of the Stop Buffer, pH 5.5, stops the enzyme-catalyzed reaction. Equilibrated Resin, which binds to the arginine, is added to the sample reactions and the reactions are then pipetted into spin cups. The citrulline, being ionically neutral at pH 5.5, flows through the cups completely. NOS activity is then quantitated by counting the radioactivity in the eluate.

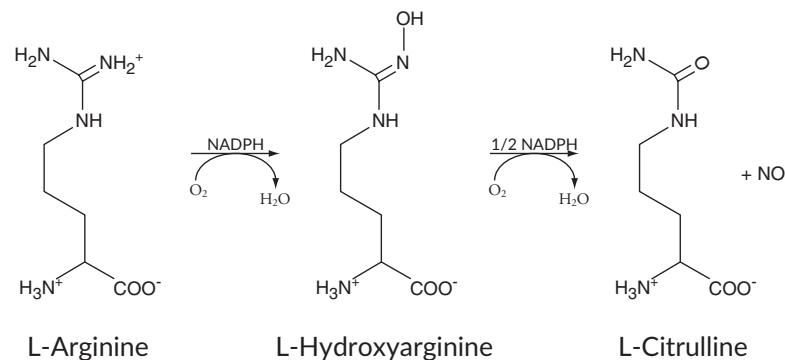


Figure 1. NOS catalyzes a 5-electron oxidation of a guanidino nitrogen of L-arginine to generate NO and L-citrulline. L-Hydroxyarginine is formed as an intermediate that is tightly bound to the enzyme. Both steps in the reaction are dependent on calcium and calmodulin.

Sample Preparation

Preparation of Extracts from Tissues and Cultured Cells

The citrulline assay has been used to quantitate levels of NOS activity in tissue homogenates from numerous sources including blood vessels, immune cells, visceral organs, nervous tissue, and cultured cells. NOS enzymes are relatively unstable; therefore, tissues should be harvested quickly after animal euthanasia. If enzyme assays are to be conducted at a later time, it is best to freeze intact tissues or harvested cultured cells prior to homogenization. Wrap the tissues in aluminum foil, flash freeze the tissues in liquid nitrogen, and then store at -80°C.

Extraction of Proteins from Tissues

NOTE: Because the level of NOS activity will vary between tissues, the volume of 1X Homogenization Buffer used may require optimization.

1. Prepare an appropriate volume of Homogenization Buffer (1X) (i.e., a 1:10 dilution of the Homogenization Buffer (10X) (Item No. 10007063). Add 10-20 volumes [volume of buffer (ml):weight of tissue (g)] of ice-cold Homogenization Buffer (1X) to a tissue sample (for tissues with low NOS activity, start with 5-10 volumes of Homogenization Buffer (1X)).
2. Homogenize the tissue using a tissue grinder or an equivalent tissue homogenizer. Keep the tissue homogenate on ice.
3. Centrifuge at 10,000 x g for 15 minutes at 4°C (or at full speed for five minutes in a microcentrifuge at 4°C).
4. Transfer the supernatant to fresh microcentrifuge tubes and keep the tubes on ice until use. This extract will be used in the NOS Activity Assay (see step 3 of **Preparing the Reactions**, page 13).

Subcellular Tissue Distribution

The subcellular distribution of NOS is tightly regulated in tissues. eNOS is largely membrane associated as a result of N-terminal myristoylation.^{9,10} nNOS is largely soluble in adult rat brain, yet in skeletal muscle, it is predominantly associated with membrane fractions.¹¹ The mechanism for membrane attachment of nNOS remains unclear. iNOS is a soluble enzyme. NOS activity in soluble and membrane-associated fractions can be separated by centrifuging the homogenized tissues at 100,000 x g for 60 minutes. The supernatant contains soluble NOS, while the pellet, which is resuspended in Homogenization Buffer, contains membrane-associated NOS.

Extraction of Proteins from Tissue Culture Cells

Certain cultured cells, such as endothelial cells and activated macrophages, contain NOS, which can be measured using the citrulline assay. The proteins must first be extracted from the cells as follows:

1. Remove the culture media from the tissue culture cells.
2. Wash the tissue culture cells once with phosphate-buffered saline (PBS) and harvest the tissue culture cells in PBS containing 1 mM EDTA. Transfer the tissue culture cells to microcentrifuge tubes.
3. Spin the microcentrifuge tubes in a microcentrifuge at full speed for two minutes to pellet the cells.
4. Remove the supernatant from the pelleted cells by vacuum aspiration and then add 100-500 µl of the Homogenization Buffer (1X) to each microcentrifuge tube of pelleted cells. Sonicate briefly to disrupt the cells.
5. Spin the microcentrifuge tubes in a microcentrifuge at full speed for five minutes.
6. Separate the supernatant from the pellet and adjust the resulting protein sample to a concentration of 5-10 mg/ml.

Reagent Preparation

1. iNOS Positive Control - (Item No. 10007068)

The vial contains 50 μ l of murine recombinant iNOS. Thaw the enzyme on ice before use. It is then ready to use as a control in the assay.

2. Calmodulin (1 μ M) - (Item No. 10007061)

The vial contains 1 μ M calmodulin. It is ready to use in the assay.

3. Reaction Buffer (2X) - (Item No. 10007062)

The vial contains 1.25 ml of 50 mM Tris-HCl, pH 7.4, containing 6 μ M tetrahydrobiopterin (BH_4), 2 μ M flavin adenine dinucleotide, and 2 μ M flavin adenine mononucleotide.

4. Homogenization Buffer (10X) - (Item No. 10007063)

The vial contains 50 ml of 250 mM Tris-HCl, pH 7.4, containing 10 mM EDTA and 10 mM ethyleneglycol-*bis*(β -aminoethylether)-N,N,N',N'-tetraacetic acid (EGTA). See under **Sample Preparation** to determine how much Homogenization Buffer (1X) to make.

5. L-NNA (10 mM) - (Item No. 10007067)

The vial contains 100 μ l of 10 mM L-NNA. L-NNA is a non-specific NOS inhibitor. It will eliminate all of the NOS activity. It is ready to use in the assay.

6. Stop Buffer - (Item No. 10007064)

The vial contains 25 ml of 50 mM N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid (HEPES), pH 5.5, and 5 mM EDTA. It is ready to use in the assay.

7. Equilibrated Resin - (Item No. 10007065)

The vial contains 5 ml of resin. It is ready to use.

8. Calcium Chloride (6 mM) - (Item No. 10007066)

The vial contains 400 μ l of 6 mM calcium chloride. It is ready to use to assay nNOS and eNOS.

9. Spin Columns and Collection Tubes - (Item No. 601223)

Sufficient spin columns and collection tubes are provided for 50 total reactions.

Performing the Assay

Measurement of Nitric Oxide Synthase Activity in Enzyme Extracts

NOTE: Refer to Appendix (on page 16) for notes on the stability of radiolabeled arginine.

Incubation of the citrulline assay reaction may be carried out for 10-60 minutes at 22-37°C depending on the tissue being used. High levels of nNOS in nervous tissues¹² and skeletal muscle¹¹ permit brief assays (10-15 minutes) of NOS with room temperature incubations. Lower levels of eNOS in vascular tissues require that assays be performed for prolonged periods (60 minutes) at room temperature.

eNOS and nNOS require calcium for enzyme activity; therefore, it is essential to add calcium to experimental assays. A final free calcium concentration of 75 μM is required for optimal NOS activity.¹² When testing NOS activity from tissue extracts, addition of calmodulin to the reaction is not required. However, when testing purified NOS, the addition of calmodulin is required for nNOS and eNOS (for review, see Reference 1, page 18). Crude extracts of macrophage iNOS, such as the 100,000 x g supernatant solution, require 1 mM of Mg^{2+} for maximum activity, while purified iNOS shows no such dependence.

NOS activity in the citrulline assay is defined as counts per minute (cpm) in an incubated test sample as compared to an appropriate blank. The following control reactions can serve as a blank: a reaction that includes 1 mM N^{G} -nitro-L-arginine (L-NNA, a competitive NOS inhibitor provided at a concentration of 10 mM), a reaction in which the extract is boiled prior to the assay, a reaction in which either NADPH or calcium (for nNOS and eNOS) is omitted, or a reaction that is incubated on ice. As for any quantitative enzyme assay, it is important to verify reaction conditions are such that the assay is linear with respect to time and protein concentration. Specific activity and substrate affinity of NOS can be assessed by carrying out replicate reactions in the presence of varying amounts of unlabeled arginine. The K_m (Michaelis constant) of NOS is in the range of 2-20 μM .^{7,10,13} Appropriate concentrations of arginine for kinetic studies are 0.1-100 μM .

Preparing the Reactions

1. Prepare and store the Reaction Mixture on ice by adding the following components to a microcentrifuge tube (500 μl or 1.5 ml):

NOTE: The volumes given here yield sufficient Reaction Mixture for 10 reactions. The Reaction Mixture can be stored on ice for up to 24 hours. Inducible NOS (iNOS) is Ca^{2+} independent. When assaying iNOS, substitute 50 μl HPLC-grade water for CaCl_2 or add 50 μl of 8 mM MgCl_2 (see page 12).

250 μl of Reaction Buffer (2X)

50 μl of 10 mM NADPH (freshly prepared in 10 mM Tris-HCl, pH 7.4)

10 μl of [³H]arginine (1 $\mu\text{Ci}/\mu\text{l}$) -or- 5 μl [¹⁴C]arginine (100 $\mu\text{Ci}/\text{ml}$)

50 μl of CaCl_2 -or- 50 μl of HPLC-grade water -or- 50 μl MgCl_2

40 μl of HPLC-grade water for [³H]arginine -or- 45 μl of HPLC-grade water for [¹⁴C]arginine

2. Label 1.5 ml microcentrifuge tubes for the appropriate reactions below.
3. **iNOS Positive Control** - add 40 μl of Reaction Mixture, 5 μl of HPLC-grade water, and 5 μl of iNOS Positive Control to a microcentrifuge tube.
4. **Control Sample** - add 40 μl of Reaction Mixture, 5 μl of calmodulin*, and 5 μl of tissue extract or NOS enzyme preparation to a microcentrifuge tube.
5. **Inhibition of Control Sample (optional)** - To chemically inhibit all of the NOS activity in the control sample (background), add 5 μl of L-NNA to 40 μl of Reaction Mixture before adding the 5 μl of tissue extract or NOS enzyme preparation to a microcentrifuge tube. If needed, add 5 μl of calmodulin* to the mixture.

6. Incubate the reactions at 22-37°C for 10-60 minutes. (For initial experiments, the reaction should be allowed to proceed at room temperature for 30 minutes.)
7. Stop the reactions by adding 400 µl of Stop Buffer to each microcentrifuge tube.

*nNOS and eNOS require 0.1 µM calmodulin when assayed as purified enzymes. The addition of calmodulin to tissue extracts is not necessary, but it is recommended. If the addition of calmodulin is required, add calmodulin to a final concentration of 0.1 µM (*i.e.*, add 5 µl to a 50 µl reaction).

Processing the Samples

1. Thoroughly resuspend the Equilibrated Resin provided. Pipette 100 µl of the Equilibrated Resin into each reaction sample.
2. Place the spin column into collection tubes and transfer the reaction samples to spin columns.
3. Centrifuge the spin columns and collection tubes in a microcentrifuge at full speed for 30 seconds.
4. Remove the spin columns and collection tubes and transfer the eluate (*i.e.*, the “flowthrough”) to scintillation vials. Add scintillation fluid to the vials and quantitate the radioactivity in a liquid scintillation counter.

Additional Controls

1. Total Counts

To determine total counts used in the reaction, transfer 40 µl of the Reaction Mixture to a scintillation vial. Add 400 µl of Stop Buffer. Add scintillation fluid and quantify the radioactivity.

2. Background Counts

To determine background counts used in the reaction, transfer 40 µl of the Reaction Mixture to a microcentrifuge tube. Add 400 µl of Stop Buffer and 100 µl of Equilibrated Resin. Process the Control the same as described for the samples (*i.e.*, apply to spin columns and count the eluate).

ANALYSIS

Calculations

The percent citrulline formed in the reaction in relation to total possible counts can be determined as follows:

$$\% \text{ Conversion} = [(\text{cpm reaction} - \text{cpm background}^*) / \text{cpm total}] \times 100$$

*The cpm from the “blank” as described on page 12 can also be used here.

In the tissues assayed according to these protocols, citrulline is the major radiolabeled compound in the eluate. This can be readily verified using thin-layer chromatography (TLC). Separation of arginine and relevant metabolites is achieved using silica-gel chromatography plates developed with CH₃OH:NH₄OH (6:1). With this solvent system, arginine migrates at an R_f (the ratio of the distance traveled by a compound to the distance traveled by the solvent) of ~0.1 while citrulline migrates at an R_f of ~0.5.

Appendix

Stability of Radiolabeled Arginine

Prior to initiating the enzyme assays, it is essential to verify the purity of the radiolabeled arginine, otherwise a high blank value for the liquid scintillation counting will greatly reduce the sensitivity of the assay. To assess the blank value, a Reaction Mixture is applied to the Equilibrated Resin. Nonadherent radioactivity is eluted with Stop Buffer, the eluate is collected and the radioactivity is quantitated in a liquid scintillation counter.

1. Prepare a reaction mix by combining the following components in a microcentrifuge tube.
 - 20 μ l of Reaction Buffer (2X)
 - 4 μ l of 10 mM NADPH [freshly prepared in 10 mM Tris, pH 7.4]
 - 1-10 μ l of [3 H]arginine (1 μ Ci/ μ l) or [14 C]arginine (100 μ Ci/ml)
 - dH₂O to bring the total volume to 40 μ l
2. Store the reaction mix on ice.
3. Perform the control experiments (*i.e.*, total counts and background counts) as described above using 10 μ l of the Reaction Cocktail.

Greater than 90% of the applied radioactivity should be retained by the spin cup. This represents a relatively low blank value. If more than 10% of the radioactivity flows through the spin cup, it is important to purchase new arginine prior to conducting the assay. [3 H]Arginine is prone to radiolytic decay and may degrade significantly within two months; [14 C]arginine is more stable but more expensive.

Quick Reference Protocol

Preparation of Extracts from Tissues and Cultured Cells

Extraction of Proteins from Tissues

- Add 10-20 volumes of ice-cold Homogenization Buffer (1X) to a tissue sample.
- Homogenize the tissue and spin the homogenate in a microcentrifuge for five minutes at 4°C (or 10,000 x g for 15 minutes at 4°C).
- Transfer the supernatant to a fresh tube and keep the tube on ice until use.

Extraction of Proteins from Tissue Culture Cells

- Remove the culture media from the tissue culture cells, wash the cells with PBS and harvest the cells in PBS containing 1 mM EDTA.
- Spin the cells at full speed for two minutes, remove the supernatant by vacuum aspiration and resuspend the cells in Homogenization Buffer.
- Sonicate or homogenize the cells.
- Spin the cells at full speed in a microcentrifuge for five minutes at 4°C, remove the supernatant, and adjust the protein sample to a concentration of 5-10 mg/ml.

Measurement of Nitric Oxide Synthase Activity in Enzyme Extracts

- Prepare the Reaction Mixture in a microcentrifuge tube and store on ice.
- Add the cell/tissue extract to 40 μ l of the Reaction Mixture.
- Incubate the reaction at 22-37°C for 10-60 minutes.
- Add 400 μ l of Stop Buffer and 100 μ l of Equilibrated Resin to the reaction sample.
- Transfer the reaction sample to a spin column and spin at full speed for 30 seconds.
- Transfer the eluate to a scintillation vial, add scintillation fluid, and quantitate the radioactivity in a liquid scintillation counter.

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NOTES

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Buyer agrees to purchase the material subject to Cayman's Terms and Conditions. Complete Terms and Conditions including Warranty and Limitation of Liability information can be found on our website.

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